(19) World Intellectual Property Organization International Bureau





(43) International Publication Date 12 July 2001 (12.07.2001)

PCT

(10) International Publication Number WO 01/49832 A2

(51) International Patent Classification7: C12N 9/00

(21) International Application Number: PCT/EP01/00060

(22) International Filing Date: 5 January 2001 (05.01.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data: 00100351.6 7 January 2000 (07.01.2000) ER 00124595.0 10 November 2000 (10.11.2000) ER

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(81) Designated States (national): AE, AG, AL, AM, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CN, CR, CU, CZ, DM, DZ, EE, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, RO, RU, SD, SG, SI, SK, SL, TJ, TM, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

 Without international search report and to be republished upon receipt of that report.

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



49832 A2

(54) Title: TRANSDUCTION OF RECOMBINASES FOR INDUCIBLE GENE TARGETING

WO 01/49832 PCT/EP01/00060

Transduction of recombinases for inducible gene targeting

The present invention provides the use of a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain for preparing an agent for inducing target gene alteration in a living organism or in cultured cells, suitable fusion proteins and a method for the production of said fusion proteins.

Background

For some years targeted mutagenesis in totipotent mouse embryonic stem (ES) cells has been used to inactivate genes, for which cloned sequences were available (Capecchi, Trends in Genetics 5, 70 - 76 (1989)). Since ES cells can pass mutations induced in vitro to transgenic offspring *in vivo*, it is possible to analyze the consequences of gene disruption in the context of the entire organism. Thus, numerous mouse strains with functionally inactivated genes ("knock out mice") have been created by this technology and utilized to study the biological function of a variety of genes.

A refined method of targeted mutagenesis, referred to as conditional mutagenesis, employs a site-specific recombination system (e.g. Cre/loxP or Flp/frt – Sauer and Henderson, N. Proc. Natl. Acad. Sci. USA 85, 5166-5170 (1988); Senecoff et al., J. Mol. Biol., 201, 405 - 421 (1988)) which enables a temporally and/or spatially restricted alteration of target genes (Rajewsky et al., J. Clin. Invest., 98, 600 - 603 (1996)). The creation of conditional mouse mutants requires the generation of two mouse strains, i.e. the recombinase recognition strain and the recombinase expressing strain. The recombinase recognition strain is generated by homologous recombination in ES cells as described above except that the targeted

exon(s) is (are) flanked by two recombinase recognition sequences (hereinafter "RRS"; e.g. loxP or frt). The type of recombination event mediated by the recombinase depends on the disposition of the RRS, with deletions, inversions, translocations and integrations being possible (Torres and Kühn, Oxford University Press, Oxford, New York (1997)). By placing the RRS into introns, an interference with gene expression before recombination can be avoided. The recombinase expressing strain contains a recombinase transgene (e.g. Cre, Flp) whose expression is either restricted to certain cells and tissues or is inducible by external Crossing of the recombinase recognition strain with the agents. recombinase expressing strain recombines the RRS-flanked exons from the doubly transgenic offspring in a prespecified temporally and/or spatially restricted manner. Thus, the method allows the temporal analysis of gene function in particular cells and tissues of otherwise widely expressed genes. Moreover, it enables the analysis of gene function in the adult organism by circumventing embryonic lethality which is frequently the consequence of gene mutation. For pharmaceutical research, aiming to validate the utility of genes and their products as targets for drug development, inducible mutations provide an excellent genetic tool. However, the current systems for inducible recombinase expression in transgenic animals suffer from a certain degree of leakiness in the absence of the inducer (Kühn et al., Science 269(5229):1427-9 (1995); Schwenk et al., Nucleic Acids Res.; 26(6):1427-32 (1998)). Furthermore, the generation of conditional mutants is a time consuming and labor intensive procedure, since the recombinase recognition strain and the recombinase expressing strain have to be breed at least over two generations in order to obtain animals carrying both, the recombinase transgene and two copies of the RRS-flanked target gene sequence.

Protein tranduction domains (hereinafter shortly referred to as "PTD") that have the ability to cross cell membranes were identified, e.g. in the

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Antennapedia protein from *Drosophila* (Vives et al., J. Biol. Chem, 272(25):16010-7 (1997)), Kaposi fibroblast growth factor (Kaposi FGF; Lin et al., J. Biol. Chem. 270: 14255-58 (1995)), VP22 from HSV (Elliott and O'Hare, Cell, 88(2):223-33 (1997)) and TAT from HIV (Green and Loewenstein, Cell, 55(6):1179-88 (1988); Frankel and Pabo, Cell, 55(6):1189-93 (1988)). WO 99/29721 moreover mentions TAT mutants having an enhanced activity as compared to the wild-type peptide.

Fusion of PTDs to heterologuous proteins conferred the ability to transduce into cultured cells (Fawell et al., Proc. Natl. Acad. Sci. USA, 91(2):664-8 (1994); Elliott and O'Hare (1997), Phelan et al., Nature Biotech. 16; 440-443 (1998) and Dilber et al., Gene Ther., 6(1):12-21 (1999)). Dalby and Bennett showed that a fusion protein consisting of VP22 and functional Flp recombinase translocated between cells in culture (from COS-1 cells transfected with VP22-Flp to CHO cells carrying Flp recognition sites (FRT sites); see Dalby and Bennett, Invitrogen, Expressions 6.2, page 13 (1999)). Further WO 99/11809 mentions a fusion protein Antp-Cre and emphasizes that it may be used to deliver the Cre into the cell which recombines inside the cell nucleus. It is mentioned that the fusion protein is suitable for manipulating genomic DNA at precise locations in a temporal regulated manner.

Furthermore, a recent report demonstrated that the ß-galactosidase protein fused to the 11 amino acids PTD from the HIV TAT protein can infiltrate all tissues of living mice reaching every single cell (Schwarze et al., Science, 285(5433):1569-72 (1999)). Finally, WO 99/60142 discloses vector constructs for gene therapy carrying a tumor cell sensitizing gene, a sensitizing gene expression regulatory system, a control gene and a control gene expression regulatory system, wherein the control gene can be a fusion gene consisting of a recombinase (viz. Cre or Flp) and a trafficking protein (viz. VP22).

With regard to the fusion protein Antp-Cre of WO 99/11809, it is however, general knowledge in the art that the Antennapedia PTD is not a generally applicable transducing protein, namely it has only a limited activity with proteins having more than 100 amino acid residues (Derossi et al., Trends Cell Biol. 8: 84-87, 1998). In view of the limited transducing activity of the Antp PTD and the size of the generally known recombinases (ranging from about 200 to about 600 amino acid residues), it was desirable to provide a more potent system for the transduction of recombinases. It was, however, not clear for a person skilled in the art whether PTDs would be effective at all with recombinases for the following reasons: (i) only a single example of PTD-mediated delivery of proteins (above 100 amino acid residues) in vivo has been reported so far (Schwarze et al., Science, 285(5433):1569-72 (1999); Fawell et al., PNAS, 91: 664-68 (1994); both references describing the TAT-mediated transduction of $\ensuremath{\mbox{\mbox{\footnotesize B-}}}$ galactosidase in mice);

- (ii) it is known that due to defolding and refolding processes the transduction of native proteins into cells may result in a significant loss of protein activity (e.g., as described for TAT-GFP; Schwarze et al, Trends Cell Biol. 10: 290-95 (2000));
- (iii) neither the number of protein molecules that can be transferred into a cell by a given translocation domain has been systematically determined, nor the number of Cre molecules in the cell nucleus that is required for efficient recombination;
- (iv) the delivery of active proteins requires unfolding- and proper refolding which is unpredictable for a given protein (Bonifaci et al., AIDS 9: 995-1000 1995); and
- (v) the mechanism by which protein transduction domains facilitate protein transduction in unknown and several findings have been published that rule out classical receptor-, transporter-, endosome- or endocytosis-mediated processes in the transduction of Ant, TAT and VP22 (G. Eliott, P. O'Hare, Cell 88, 223-233 (1997); D.A. Mann, A.D. Frankel, EMBO. J. 10,

1733-1739 (1991); D. Derossi et al., J. Biol. Chem. 269, 10444-10450 (1994); D. Derossi et al., J. Biol. Chem. 271, 18188-18193 (1996); E. Vives et al., J. Biol. Chem. 272, 16010-16017 (1997)).

Moreover, there was still the need for a generally applicable method where the genetic manipulation can be performed in both, endogenous genes and transgenes.

Summary of the Invention

It was found that site-specific DNA recombinase proteins can be translocated into cells of a living organism when fused to specific protein transduction domains, namely transduction domains being derived from the VP22 protein of HSV or from the TAT protein of HIV. Thus, whenever a gene mutation is desired, recombination is induced upon the injection of the appropriate site-specific recombinase fused to a transduction domain into such a living organism (provided, however, that said organism carries at least one appropriate RRS integrated in the genome).

The present invention thus provides

- (1) the use of a fusion protein comprising
- (a) a site-specific DNA recombinase domain and
- (b) a protein transduction domain (PTD)

for preparing an agent for inducing target gene alterations in a living organism or cell culture, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome;

(2) a method for inducing gene alterations in a living organism which comprises administering to said living organism a fusion protein comprising a site-specific DNA recombinase domain and a PTD as defined in (1) above, wherein said living organism carries at least one or more

recognition sites for said site-specific DNA recombinase integrated in its [] genome;

- (3) a fusion protein comprising
- (a) a site-specific DNA recombinase domain and
- (b) a PTD being derived from the VP22 protein of HSV or from the TAT protein of HIV

provided that when the site-specific DNA recombinase domain is wild-type Cre or Flp then the PTD is not the full length VP22 PTD of HSV (i.e., the fusion protein is not identical to the fusion protein of Dalby and Bennett, Invitrogen, Expressions 6.2, page 13 (1999) and of WO 99/60142);

- (4) a DNA sequence coding for the fusion protein of (3) above;
- (5) a vector comprising the DNA sequence as defined in (4) above;
- (6) a host cell transformed with the vector of (5) above and/or comprising the DNA of (4) above;
- (7) a method for producing the fusion protein of (1) above which comprises culturing the transformed host cell of (6) above and isolating the fusion protein; and
- (8) an injectable composition comprising the fusion protein as defined in (1) or (3) above.

The invention is further illustrated by the appended Figures and is explained in detail below.

Description of the Figures

<u>Fig. 1:</u> Generation of induced mouse mutants using purified fusion proteins.

A: Expression of the fusion protein consisting of the site-specific DNA recombinase (e.g. Cre) and the protein transduction domain (e.g. the HIV derived TAT peptide) in prokaryotic or eukaryotic cells.

B: Extraction and purification of the expressed fusion protein (e.g. as described in Nagahara et al., Nat. Med. 4 (12):1449-52 (1998)).

C: Injection of the purified fusion protein into mice carrying the RRSflanked target sequence.

D: Analysis of the pattern of induced target gene recombination and the resulting phenotype.

Triangle: RRS.

Fig. 2: Scheme of the bacterial expression vector pT7-TACS (SEQ ID NO:16). The coding region of the 11 amino acid protein transduction domain of HIV TAT protein is fused to the N-terminus of the Cre recombinase protein sequence. The 10-amino-acid strep tag and the protease factor Xa recognition sequence are fused to the C-terminus. The T7 promoter permits expression of TAT-Cre protein in *E. coli*.

<u>Fig. 3:</u> Detection of purified TAT-Cre protein by Coomassie staining and Western blot analysis.

A: Coomassie stained SDS-PAGE gel. Lane 1: 10 kDa ladder (Life Technologies, Cat. No.: 10064-012), 2: 1000 ng BSA, 3: 750 ng BSA, 4: 500 ng BSA, 5: 100 ng BSA, 6: 50 ng BSA, 7: 5 µl TAT-Cre, 8: 1 µl TAT-Cre in Bicine buffer.

B: Western blot analysis using an alkaline phosphatase-conjugated antistrep tag antibody (IBA, Cat. No: 2-1503-001). Lane 1: MultiMark (Invitrogen, Cat. No.: LC5725), 2: 7 µl TAT-Cre, 3: 5 µl TAT-Cre, 4: 2,5 µl TAT-Cre, 5: 1,25 µl TAT-Cre in Bicine buffer.

Fig. 4: X-Gal staining of M5Pax8 cells treated with TAT-Cre protein. M5Pax8 fibroblasts where treated for 18 h with 3,5 (A), 6,9 (B) and 13,8 μg/ml TAT-Cre protein (C) in serum-free medium. Four days after treatment, cells were fixed and stained with X-Gal.

<u>Fig. 5:</u> Measurement of β-galactosidase activity in cell lysates. M5Pax8 fibroblasts where treated for 18 h with increasing concentrations of TAT-Cre, as indicated, or transiently transfected with either expression vectors

for Cre (pCMV-I-Cre-pA, see SEQ ID NO:29) or β-galactosidase (pCMV-I-β-pA, see SEQ ID NO:30). Four days after treatment, cells were lysed and the β-galactosidase activities were determined.

Fig. 6: PCR detection of TAT-Cre mediated recombination in mice.

A: PCR-analysis of genomic DNA from duodenum (lane 2), liver (3), kidney (4), spleen (5), muscle (6), lung (7), tail (8) and brain (9) of a pln13 mouse treated three times with intraperitoneal injections of 75 μg TAT Cre protein at two-day-intervals. Deletion of the loxP-flanked DNA segment is indicated by the presence of the about 400 bp fragment. Lane 1: 1-kb-ladder (Life Technologies).

B: PCR strategy to detect Cre-mediated deletion of the loxP-flanked DNA segment. Arrows indicate the positions of the primers.

C: PCR-analysis of genomic DNA from spleen of a *pln13* mouse treated three times with intraperitoneal injections of 75 µg TAT Cre protein at two-day-intervals (lane 4). To confirm the presence of the BamH I restriction site, the PCR product was digested with BamH I which produces two diagnostic fragments of about 190 and about 210 bp (5). As a control, tail DNA from untreated mice carrying the loxP-flanked (lane 2) and the detected pln13 allele (3) was subjected to PCR amplification. Lane 1: 100 bp ladder (Life Technologies), lane 6: 1 kb ladder (Life Technologies).

Fig. 7: Scheme of the bacterial expression vectors pT7-VPCS (SEQ ID NO:17) and pCRT7- Δ VPCS (SEQ ID NO:15). The coding region of the 301 amino acid protein transduction domain of HSV VP22 protein (A) or the truncated 143 amino acid Δ VP22 domain (B) is fused to the N-terminus of the Cre recombinase protein sequence. The 10-amino-acid strep tag and the protease factor Xa recognition sequence are fused to the C-terminus. The T7 promoter allows the expression of VP22-Cre and Δ VP22-Cre fusion proteins in *E. coli*. The sequence in pCRT7- Δ VPCS encoding the 15 amino

acid N-terminal leader sequence is used for enhanced protein stability (Invitrogen).

<u>Fig. 8:</u> Detection of the purified VP22-Cre and Δ VP22-Cre fusion proteins by Coomassie staining and Western blot analysis.

A: Detection of VP22-Cre protein in a Coomassie-stained SDS-PAGE gel.
Lane 1: 10 kDa ladder, 2: 1000 ng BSA, 3: 500 ng BSA, 4: 100 ng BSA,
5: inclusion body protein extract before chromatography, 6: unbound
protein, 7: fraction 17, 8: fraction 18, 9: fraction 19, 10: fraction 20. The
position of the 75 kDa VP22-Cre protein is indicated by the arrow head.
B: Detection of VP22-Cre protein by Western blot analysis using an
alkaline phosphatase-conjugated anti-strep tag antibody (IBA, Cat. No.: 21503-001). Lane 1: MultiMark (Invitrogen), 2: inclusion body protein
extract before chromatography, 3: unbound protein, 4: fraction 10, 5:
fraction 11, 5: fraction 16, 6: fraction 17, 7: fraction 18, 8: fraction 19, 9:
fraction 19, 10: fraction 20.

C: Detection of Δ VP22-Cre protein in a Coomassie-stained SDS-PAGE gel. Lane 1: 10 kDa ladder, 2: inclusion body protein extract before chromatography, 3: unbound protein, 4: fraction 1, 5: fraction 8, 6: fraction 9, 7: fraction 15, 8: 100 ng BSA, 9: 500 ng BSA, 10: 1000 ng BSA. The position of the 60 kDa Δ VP22-Cre protein is indicated by the arrow head.

D: Detection of \triangle VP22-Cre protein by Western blot analysis using a alkaline phosphatase-conjugated anti-strep tag antibody (IBA, Cat. No.: 2-1503-001). Lane 1: MultiMark (Invitrogen), 2: inclusion body protein extract before chromatography, 3: unbound protein, 4: fraction 4, 5: fraction 8, 6: fraction 10, 7: fraction 12, 8: soluble protein extract before chromatography, 9: unbound protein, 10: fraction 7.

<u>Fig. 9:</u> X-Gal staining of M5Pax8 cells treated with VP22-Cre and Δ VP22-Cre fusion proteins. M5Pax8 fibroblasts where treated for 18 h with either

Bicine buffer (A), 0.5 μ g/ml VP22-Cre (B) or 3.75 g/ml Δ VP22-Cre (C) in serum-free medium. Four days after treatment, cells were fixed and stained with X-Gal.

<u>Fig. 10:</u> Measurement of β-galactosidase activity in cell lysates. M5Pax8 fibroblasts where treated for 18 h with VP22-Cre, ΔVP22-Cre or Bicine buffer alone, as indicated or transiently transfected with expression vectors for Cre (pCMV-I-Cre-pA, see SEQ ID NO:29) or β-galactosidase (pCMV-I-β-pA, see SEQ ID NO:30). Four days after treatment, cells were lysed and the β-galactosidase activities were determined.

<u>Fig. 11:</u> PCR detection of Cre mediated recombination in cells treated with VP22-Cre and Δ VP22-Cre fusion proteins shown in SEQ ID NOs: 21 and 14, respectively).

A: PCR-analysis of genomic DNA isolated from M5Pax8 fibroblasts. Cells were transiently transfected with a Cre expression vector (lane 2) or treated for 18 h with either buffer alone (lane 3), 7.5 μ g/ml VP22-Cre (4, 5) or 15 μ g/ml Δ VP22-Cre (6, 7) in serum-free medium. Four days after treatment, genomic DNA was extracted and subjected to PCR amplification. Deletion of the loxP-flanked DNA segment is indicated by the presence of the 226 bp DNA fragment. To confirm the presence of the Nco I restriction site in the recombined allele, the PCR products were digested with Nco I which produces two diagnostic fragments of 85bp and 141bp (lanes 5 and 7). Lane 1: 100 bp ladder (Life Technologies), lane 8: 1 kb ladder (Life Technologies).

B: PCR strategy to detect Cre-mediated deletion of the loxP-flanked DNA segment. Arrows indicate the positions of the primers.

Detailed Description of the Invention

The expression "target sequences" according to the present invention means all kind of sequences which may be mutated (viz. deleted,

WO 01/49832 PCT/EP01/00060

11

translocated, integrated and/or inverted) by the action of the recombinase. The number of RRS in the target sequence depends on the kind of mutation to be performed by the recombinase. For most of the mutations (especially for deletions and invertions) two RRS are required which are flanking the sequence to be mutated (deleted or inverted). For some kinds of integrations only one RRS may be necessary within the target sequence.

The "living organisms" according to the present invention are multi-cell organisms and can be vertebrates such as mammals (e.g., rodents such as mice or rats) or non-mammals (e.g., fish) or can be invertebrates such as insects or worms, or can be plants (higher plants, algi or fungi). Most preferred living organisms are mice and fish.

"Cell culture" according to the present invention include cells isolated from the above defined living organism and cultured *in vitro*. These cells can be transformed (immortalized) or untransformed (directly derived from the living organism; primary cell culture).

The site-specific DNA recombinase domain within the fusion protein of the invention of the present application is preferably selected from a recombinase protein derived from Cre, Flp, ϕ C31 recombinase (Thorpe and Smith, Proc. Natl. Acad. Sci, USA, vol. 95, 5505-5510 (1998)), $\gamma\delta$ resolvase (Schwickardi and Dröge, FEBS letters 471:147-150 (2000) and R recombinase (Araki et al., J. Mol. Biol., 182, 191-203 (1985)). The preferred recombinases are Cre and mutants thereof (preferably the Cre variant of aa 15 to 357 of SEQ ID NO: 2 or aa 325-667 of SEQ ID NO: 6) and Flp and variants thereof including Flpe (preferably the Flp variant of aa 15 to 437 of SEQ ID NO: 4 or aa 325 to 747 of SEQ ID NO: 8).

PCT/EP01/00060

The protein transduction domain according to the present invention includes, but is not limited to, the PTDs mentioned in Background of the Invention. The PTD preferably is derived from the VP22 protein of HSV or from the TAT protein of HIV. Suitable TAT proteins include, but are not limited to, proteins comprising (i) the amino acid sequence shown in SEQ

ID NO: 10 and mutant thereof such as

(ii) proteins comprising the amino acid

AGRKKRRQRRR (SEQ ID NO:22)

YARKARRQARR (SEQ ID NO:23)

YARAAARQARA (SEQ ID NO:24)

YARAARRAARR (SEQ ID NO:25)

YARAARRAARA (SEQ ID NO:26)

YARRRRRRRR (SEQ ID NO:27)

YAAARRRRRR (SEQ ID NO:28)

as known from WO 99/29721. Preferred are transduction domains consisting of the TAT proteins (i) and (ii) above.

Suitable VP22 proteins include, but are not limited to, the wild-type VP22 protein, i.e., a protein comprising amino acids 1 to 302 of SEQ ID No:21, and truncated forms thereof. Truncated VP22 proteins in accordance with the present invention can be those lacking 1 to 158 amino acid residues at their N-terminal end. The most preferred VP22 protein is the truncated VP22 PTD comprising amino acid residues 16 to 157 of SEQ ID NO:14.

The fusion of the two domains of the fusion protein can occur at any possible position, i.e., the protein transduction domain can be fused to the N- or C-terminal of the site-specific DNA recombinase or can be fused to active sites within the site-specific DNA recombinase. Preferably the protein transfusion domain is fused to the N-terminal of the site-specific DNA recombinase domain.

The protein transduction domain can be fused to the site-specific DNA recombinase either through a direct chemical bond or through a linker molecule. Such linker molecule can be any bivalent chemical structure capable of linking the two domains. The preferred linker molecule according to the present invention is a short peptide, e.g., having 1 to 20, preferably 1 to 10, amino acid residues. Specifically preferred short peptides are essentially consisting of Gly, Ala and/or Leu.

The fusion protein of the invention of the present application may further comprise other functional sequences such as secretion conferring signals, nuclear localisation signals and/or signals conferring protein stabilisation.

Such a preferred DNA sequence is for instance shown in SEQ ID NO: 11. In said sequence the 3' terminal codon ggc codes for the linker Gly. The DNA sequence of a suitable recombinase may be directly attached to said codon ggc.

The fusion protein can be obtained by the following steps:

 Fusion of the recombinase coding region (e.g. encoding Cre: see amino acids 15 to 357 of SEQ ID NO: 2) with the sequence conferring protein translocation (e.g. the sequence encoding the TAT peptide YGRKKRRQRRR, SEQ ID NO: 10) using standard cloning protocols (Maniatis et al., Cold Spring Harbor Laboratory, New York (1989)) or chemical synthesis. ...

- Generation of a construct for the expression of the fusion protein in prokaryotic or eukaryotic cells, e.g. in E. coli DH5a (Hanahan, J. Mol. Biol.;166(4):557-80 (1983)) using the QIAexpress pQE vector (Qiagen, Hilden).
- 3. Expression of the above mentioned fusion protein in prokaryotic or eukaryotic cells, e.g. in E. coli DH5a (Hanahan, 1983)
- 4. Extraction and purification of the above mentioned fusion protein e.g. as described in Nagahara et al., Nat. Med., 4(12):1449-52 (1998).

In an experiment it was shown that TAT-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting both in cell lines and living organisms. In this experiment a vector for the expression of a TAT-Cre fusion protein in E. coli was constructed, TAT-Cre protein was expressed in E. coli and purified from bacterial lysates. To test the activity of the TAT-Cre protein *in vitro*, a reporter cell line that contains a loxP-containing reporter construct was used. This reporter, when recombined by Cre recombinase, allows the expression of a B-galacosidase gene. Further, a transgenic mouse strain carrying a loxP-flanked target was used to invest the activity of the TAT-Cre protein *in vivo*.

In a second experiment it was shown that VP22-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting. In this experiment Bacterial expression vectors were constructed for the production of VP22-Cre fusion proteins in E. coli. The activity of purified VP22-Cre proteins were tested using a reporter fibroblast cell line containing a loxP-flanked reporter construct.

Thus, the injection of the purified fusion protein of the present invention into a living organism (e.g., a mouse) carrying a gene comprising the RRS-flanked target sequence (e.g., in an amount of 1 to 200, preferably 5

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1

to 50 µg per g body weight). To demonstrate the feasibility of the invention, a reporter mouse strain carrying an RRS-flanked cassette was used (Thorey et al., Mol. Cell Biol., 18(10):6164 (1998)).

Analysis is achieved by determining the pattern of induced target gene recombination (e.g. through PCR analysis, Southern blot analysis or X-Gal staining on tissue sections; Maniatis et al., 1989; Gossler and Zachgo, Joyner AL (Ed.), Oxford University Press, Oxford, New York (1993)).

The procedure's advantages over current technology are as follows:

- (i) The absence of background recombination before administration of the fusion protein.
- (ii) The reduction of time and resources which are necessary to combine the recombinase transgene and two copies of the RRS-flanked target gene by conventional breeding.

In experiments it was shown the following: (a) With a suitable vector for the expression of a TAT-Cre fusion protein, a TAT-Cre fusion protein was expressed in *E. coli* and purified from bacterial lysates.

- (b) A reporter cell line containing a loxP-containing reporter construct was used to test the activity of the TAT-Cre protein *in vitro*. This reporter, when recombined by Cre recombinase, allows the expression of a ß-galacosidase gene.
- (c) A transgenic mouse strain carrying a loxP-flanked target was used to invest the activity of the TAT-Cre protein *in vivo*.

These experiments demonstrate that TAT-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting both in cell lines and living organisms.

Furthermore, bacterial expression vectors were constructed for the production of VP22-Cre fusion proteins in E. coli. The activity of purified VP22-Cre proteins were tested using a reporter fibroblast cell line containing a loxP-flanked reporter construct. These experiments demonstrate that VP22-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting.

The invention is further illustrated by the following, non-limitative examples.

Examples

ii

Materials and Methods

Construction of pT7-TACS: The TAT-Cre coding region was generated by PCR using Advantage-HF PCR Kit (Clontech), 20 pmol of the primers gcg gca tgt cca att tac tga ccg tac acc-3'; SEQ ID NO:31) and TATcre antisense (5'-ttt cgg atc cgc cgc ata acc agt q-3'; SEO ID NO:32) and 10 ng pCMV-I-Cre-pA (see SEQ ID NO:29) as template. The PCR reaction was performed using the following cycle profile: 2' 94 °C, 4 x (30" 94 °C min, 30" 50 °C, 1' 72 °C), 12 x (30" 94 °C min, 30" 55 °C, 1' 72 °C) and 10' 72 °C. The resulting PCR fragment was digested with Nco I and BamH I, treated with Klenow enzyme and ligated into the plasmid pBSII KS+ which had been opened with restriction enzyme BamH I, treated with Klenow and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pBS TAT-5'cre was verified by DNA sequencing. The Plasmid pCMV-I-Cre-pA (SEQ ID NO:29) was digested with Age I and Sal I which released a 1,036 kb fragment containing the 3' part of the Cre coding region. This fragment was ligated into the plasmid pBS TAT-5'cre which had been opened with Age I and Sal I.

10 ng pBS-TATCre was subjected to PCR amplification using 20 pmol of primers FPA001 (5'-tat atc tag acc atg ggc tac ggc cgc aag aag c-3'; SEO ID NO:33) and FPA002 (5'-gct acc acg acc ttc gat acc atc gcc atc ttc cag cag gcg c-3'; SEQ ID NO:34). PCR was performed using 2,5 U Platinum Pfx DNA polymerase (Gibco BRL) and 2 x Enhancer Solution (Gibco BRL) according to the manufacturers protocol. The following cycle profile was used: 2' 94 °C, 25 x (30" 94 °C min, 15" 54,6 °C, 2'30" 68 °C). The amplified PCR fragment was purified using GFX columns (Amersham Pharmacia), digested with Xba I and ligated into the plasmid pASK57 (Skerra and Arne, Gene 151: 131-135 (1994)) which had been opened with restriction enzymes Xba I and Eco 47 III and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pASK75-TACS was digested with restriction enzymes Nco I and Hind III which released a 1,1 kb fragment. The fragment was subsequently ligated into the plasmid pT7-7 (Studier and Moffatt, J. Mol. Biol. 189: 113-130 (1986)) which had been opened with restriction enzymes Nco I and Hind III and dephosphorylated with calf intestinal phosphatase resulting in the plasmid pT7-TACS (SEQ ID NO:16).

Construction of pT7-VPCS: The Cre coding region was generated by PCR using Advantage-HF PCR Kit (Clontech), 20 pmol of the primers VP22cre sense (5'-taa cta gcg gcc gca tgt cca att tac tga ccg tac ac-3'; SEQ ID NO:35) and VP22cre antisense (5'-tcg agc ggc cgc cat cgc cat ctt cca gca ggc g-3'; SEQ ID NO:36) and 10 ng pgkcre-pA (SEQ ID NO:40) as template. The PCR reaction was performed using the following cycle profile: 2' 94 °C, 5 x (30" 94 °C, 30" 50 °C, 2' 72 °C), 15 x (30" 94 °C, 30" 55 °C, 2' 72 °C) and 10' 72 °C. The resulting PCR fragment was digested with Not I and ligated into the plasmid pVP22/Myc-His (Invitrogen), which had been opened with restriction enzyme NotI, dephosphorylated with calf intestinal phosphatase. The resulting plasmid pVP22-cre myc/His was verified by DNA sequencing.

14

10 ng pVP22-cre myc/His was subjected to PCR amplification using 20 pmol of primers FPA004 (5'-tat atc tag aca tat gac ctc tcg ccg ctc cg-3'; SEQ ID NO:37) and FPA002 (SEQ ID NO:34). PCR was performed using 2,5 U Platinum Pfx DNA polymerase (Gibco BRL) and 2 x Enhancer Solution (Gibco BRL) according to the manufacturers protocol. The following cycle profile was used: 2' 94 °C, 25 x (30" 94 °C min, 15" 54,6 °C, 2'30" 68 °C). The amplified PCR fragment was purified using GFX columns (Amersham Pharmacia), digested with Xba I and ligated into the plasmid pASK57 (Skerra and Arne, Gene 151: 131-135 (1994)) which had been opened with restriction enzymes Xba I and Eco 47 III and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pASK75-VPCS was digested with restriction enzymes Nde I and Hind III which released a 2,0 kb fragment. The fragment was subsequently ligated into the plasmid pT7-7 (Studier and Moffatt, J. Mol. Biol. 189: 113-130 (1986)) which had been opened with restriction enzymes Nde I and Hind III and dephosphorylated with calf intestinal phosphatase resulting in the plasmid pT7-VPCS (SEQ ID NO:17).

Construction of pCRT7-ΔVPCS: The ΔVP22-Cre coding region was generated by PCR using Platinum Pfx DNA polymerase (Life Technologies), 20 pmol of the primers FPA007 (5'-ttc cga aga cga cga aac acc-3'; SEQ ID NO:38) and FPA008 (5'-tat att cga agc tta tta acc acc gaa ctg cg-3'; SEQ ID NO:39) and 30 ng pT7-VPCS (SEQ ID NO:17) as template. The PCR reaction was performed using the following cycle profile: 2' 94 °C, 25 x (30" 94 °C, 30" 61 °C, 2'30" 68 °C) and 7' 68 °C. The resulting 1,8 kb PCR fragment was digested with Nco I and Sfu I and ligated into the plasmid pCRT7/VP22-1 (Invitrogen), which had been opened with restriction enzymes Nco I and Sfu I, and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pCRT7-ΔVPCS (SEQ ID NO:15) was verified by DNA sequencing.

Expression of the fusion proteins in E. coli: E. coli BL21(DE3)-RIL cells (Stratagene) were transformed with pT7-TACS and grown on LB agar plates containing 100 µg/ml ampicillin. E. coli BL21(DE3)-RP cells (Stratagene) were transformed with pT7-VPCS and grown on LB agar plates containing 100 µg/ml ampicillin. E. coli BL21(DE3)-pLysS (Invitrogen) were transformed with pCRT7-∆VPCS and grown on LB agar plates containing 25 µg/ml kanamycine and 34 µg/ml chloramphenicol. Single colonies were isolated and used to prepare glycerol stocks. Eight 5ml LB (Lura Bertani) aliquots containing antibiotics were inoculated with stabs from the glycerol stocks and grown overnight at 37°C with shaking. Two 5ml overnight cultures were each used to inoculate one of four 1L LB aliquots containing antibiotics and grown at 37°C with shaking. Growth rate was monitored by spectrophotometry at 578nm. When the cultures had obtained an $OD_{578} = 0.5$ expression of the fusion proteins were induced by the addition of 0,5 mM Isopropyl-B-D-1-thiogalactopyranosid (IPTG). Two hours after induction cells were harvested by centrifugation at 12000xg and the pellet rapidly frozen in liquid nitrogen and stored immediately at -80°C.

Purification of the fusion proteins from bacterial lysates: Each 10g cell pellet was resuspended on ice in 30ml Bicine buffer (50mM Bicine, pH 8,5) including one protease inhibitor tablet (Complete, Roche). Cells were lysed through threefold treatment (1500psi, 5 minutes) with the cell disruption bomb (Parr Instrument). 30ml of Benzonase (10000U, Merck) was added and cell extracts were incubated for 30 minutes at 4°C. Cell extracts were then centrifuged at 12,000xg (4°C). The pellet was redissolved in 8M urea, 50mM Bicine, 100mM DTT, pH 8,5 by incubation for 16 hours at 4°C. Protein extract was centrifuged at 31000xg and supernatant harvested. Protein extract was diluted in an equal volume of Chromatography buffer A (50mM Bicine, pH 8,5). PH was adjusted to pH

8,5 and the extract was filtered through a 0,45µm filter (Millipore). FPLC (Akta Explorer, Amersham Pharmacia) was performed using a cation exchange column (Sepharose SP, Column body HR_5/5 (0.5 x 5cm), column volume (CV) 1ml, linear flow 300cm/hour, Amersham Pharmacia). After addition of sample to FPLC column, buffer was exchanged with Chromatography buffer A at 10 CV.

TAT-Cre and VP22-Cre fusion proteins were eluted from the column by gradient elution using chromatography buffer B (50mM Bicine, 1M NaCl, pH 8,5) using the following profile: 0 - 50 % buffer B, 0 CV; 50 % buffer B, 10 CV; 50 - 100 % buffer B (linear gradient), 20 CV; 100 % buffer B, 10 CV. ΔVP22-Cre protein was eluted from the column by gradient elution using the following profile: 0 - 10 % buffer B, 0 CV; 10 % buffer B, 10 CV; 10 – 30 % buffer B, 0 CV; 30 % buffer B, 10 CV; 30 – 100 % buffer B, 0 CV; 100 % buffer B, 10 CV. Three 1,5ml fractions each containing purified fusion proteins were collected. Purity and concentration of protein fractions were determined by Coomassie blue stained SDS-PAGE gels and Western blot analysis using dilutions of BSA standard solutions. In addition protein content was determined using a Bradford assay (Coomassie Plus protein assay, Pierce).

SDS-PAGE and Western blot analysis: SDS-PAGE and Coomassie staining was performed according to standard protocols (Maniatis et al., Cold Spring Harbor Laboratory, New York (1989)) using 4 - 12 % gradient SDS-polyacrylamide gels (NuPAGE, Invitrogen, cat. no.: NPO321). Western blot analysis was performed using a Semi-Try Blotting Chamber (Biorad) and nitrocellulose membranes (0,2 µm; Schleicher & Schuell) according to the manufacturers protocols. The fusion proteins were detected by using an alkaline phosphatase-conjugated anti-strep tag antibody (IBA, Cat. No.: 2-1503-001) according to the manufacturers protocol.

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Generation of the M5Pax8 Cre reporter cell line: The SV40-transformed murine embryonic fibroblast line MEF5/5 (Schwenk et al., Nucl Acids Res 26(6), 1427-32 (1998)) was transfected with the vector pPGKpaX1 (Kellendonk et al, Nucl. Acids Res. 24, 1404-11 (1996)). 10^6 MEF5/5 cells were electroporated with 20 μ g pPGKpaX1 plasmid DNA linearised with Sca I and plated into 48-well-plates. The cells were cultured in DMEM/Glutamax medium (Life Technologies) supplemented with 10 % fetal calf serum at 37° C, 10 % CO_2 in humid atmosphere. Two days after transfection the medium was supplemented with 5 μ g/ml puromycine (Calbiochem) for the selection of stable integrants. 14 puromycine-resistant clones were expanded and tested by transien transfection with the Cre expression vector pPGK-Cre-pA (SEQ ID NO: 40). In two out of the 14 puromycine-resistant clones, the expression of β -galactosidase could be detected by staining with X-Gal. One of these clones, M5Pax8, was used as Cre reporter cell line.

<u>Transfection and measurement of β-galactosidase activity:</u> Fibroblasts $(10^6 \text{ cells per } 24 \text{ well plate (Falcon)})$ were transfected with 25 ng pCMV-I-Cre-pA (see SEQ ID NO:29) or pCMV-I-β-pA (see SEQ ID NO:30) plasmids using the FuGene transfection reagent (Roche Diagnostics). After 2 days the cells were lysed and the β-galactosidase activities were determined with the β-galactosidase reporter gene assay (Roche Diagnostics) according to the manufacturers guidelines using a Lumistar luminometer (MWG).

Histochemical detection of β-galactosidase activity: To quantitate β-galactosidase expression, fibroblast cells were washed once with phosphate buffered saline (PBS), and the cells were fixed for 5 minutes at room temperature in a solution of 4% formaldehyde in PBS. Next, the cells were washed twice with PBS and finally incubated in staining solution for 24 hours at 37°C (staining solution: 5 mM K3(Fe(CN)6), 5mM

K4(Fe(CN)6), 2mM MgCl2, 1mg/ml X-Gal (BioMol) in PBS). Blue stained, β-galactosidase positive cells were detected and distinguished from negative (transparent) cells in a cell culture binocular microscope under 200x magnification. For each determination a minimum of 200 cells was counted.

PCR detection of Cre-mediated recombination: Genomic DNA extracted from tissue samples was subjected to PCR using Taq-polymerase (Gibco BRL Cat. No. 10342-020) using 20 pmol of each primer (sense: 5`-CAT CTC CGG GCC TTT CGA CCT G - 3', antisense: 5'-GCG ATC GGT GCG GGC CTC TTC - 3'; SEQ ID Nos: 41 and 42, respectively). PCR was performed using the following cycle profile: 2' 94°C, 35 x (30" 94°C, 30" 55 °C, 1' 72 °C), 10 min 72 °C. PCR products were separated on a 1,2 % agarose gel.

Example 1

The vector pT7-TACS (SEQ ID NO:16) was constructed for the expression of a TAT-Cre fusion protein in E. coli. The plasmid contains the coding region of the 11 amino acid protein transduction domain of the wild-type HIV TAT protein (Green and Loewenstein, Cell, 55(6):1179-88 (1988); Frankel and Pabo, Cell, 55(6): 1189-93 (1988); SEQ ID NO:10) fused to the N-terminus of Cre recombinase protein sequence. The 10-amino-acid strep tag at the C-terminus allows the detection and purification of the fusion protein using specific antibodies (Schmidt and Skerra, J. Chromatogr A 676: 337-345 (1994)). The protease factor Xa recognition site (Ile-Glu-Gly-Arg) permits the removal of the strep tag by proteolytic cleavage. The estimated molecular weight of the TAT-Cre fusion protein is 42 kDa. A scheme of the TAT-Cre expression vector is depicted in figure 2. For the expression of TAT-Cre, the E. coli strain BL21(DE3)-RIL (Stratagene) was used. This strain carries an IPTG-inducible T7 polymerase gene and additional copies of the tRNA genes for the 'rare

codons' argU, ileY and leuW.

E. coli BL21(DE3)-RIL cells were transformed with pT7-TACS and grown in LB medium containing 100 µg/ml ampicillin. The expression of the 40 kDa TAT-Cre fusion protein could be strongly induced by the addition of 0,5 mM IPTG to the culture medium. Analysis of protein lysates revealed that approximately 50 % of TAT-Cre protein accumulated as insoluble inclusion bodies. The inclusion bodies where extracted and dissolved in 8 M urea. TAT-Cre was subsequently purified from this fraction using ion exchange chromatography. The quantity and purity of TAT-Cre protein was determined using Coomassie stained SDS-PAGE gels and Western blot analysis (figure 3). The purification process yielded TAT-Cre protein extracts of 64 % purity and a concentration of 100 µg/ml. To analyse the ability of the purified TAT-Cre protein to transduce into cultured cells, we used the fibroblast cell line M5Pax8 (R. Kühn, unpublished) that contains a loxP-containing reporter construct. This reporter, when recombined by Cre recombinase, allows the expression of a β-galacosidase gene (Buchholz et al, Nucleic Acids Res. 24, 4256-4262, 1996). Cells were cultured for 18 h with increasing concentrations of TAT-Cre protein in serum-free medium and analysed 4 days later for 6-Galacosidase activity. Staining with X-Gal showed that > 50 % of the cells treated with 13,8 μg/ml TAT-Cre protein expressed β-galactosidase indicating recombination of the loxP-flanked reporter construct had occurred (figure 4). Measurement of β-galactosidase activity in cell lysates revealed an up to 30-fold higher level of B-galactosidase activity in comparison to cells which had been transiently transfected with an eukaryotic Cre expression vector (figure 5).

To investigate the activity of TAT-Cre protein in a living organism, we used a transgenic mouse strain carrying a loxP-flanked target for Cre-mediated recombination (Thorey et al., 1998, Mol. Cell. Biol. 18: 3081-3088). Mice where treated three times with intraperitoneal injections of 75 μ g TAT Cre protein at two-day-intervals and analysed 2 days later. Genomic DNA was

isolated from a variety of organs and subjected to PCR amplification which specifically amplifies a 400 bp fragment of the recombined allele. The deleted allele could be detected in multiple tissues from treated mice indicating TAT-Cre-mediated recombination in these organs (figure 6). This experiments demonstrates that TAT-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting in cell lines and in living organisms.

Example 2

The vectors pT7-VPCS (SEQ ID NO:17) and pCRT7- Δ VPCS (SEQ ID NO:15) were constructed for the expression of VP22-Cre and Δ VP22-Cre fusion proteins in E. coli. The VP22-Cre gene of pT7-VPCS contains the full length protein translocation domain of the HSV VP22 protein (Elliott and O'Hare, Cell, 88(2): 223-33 (1987), whereas the Δ VP22-Cre gene of pCRT7- Δ VPCS contains a truncated VP22 protein transduction domain (amino acids 159 – 301; Invitrogen; aa 16-157 of SEQ ID NO:14) fused to the N-terminus of Cre recombinase protein sequence. A 10-amino-acid strep tag at the C-terminus of Cre protein sequence allows the detection and purification of the fusion proteins using specific antibodies (Schmidt and Skerra, J. Chromatogr A 676: 337-345 (1994)). The protease factor Xa recognition site permits the removal of the Strep tag by proteolytic cleavage. The estimated molecular weight is 75 kDa for VP22-Cre protein and 60 kDa for Δ VP22-Cre protein. A scheme of the vectors pT7-VPCS and pCRT7- Δ VPCS is depicted in figure 7.

E. coli BL21(DE3)-RIP cells (Stratagene) were transformed with pT7-VPCS and cultured in LB medium containing 100 μ g/ml ampicillin. E. coli BL21(DE3)-pLysS cells (Stratagene) were transformed with pCRT7- Δ VPCS and cultured in LB medium containing 25 μ g/ml kanamycine and 34 μ g/ml chloramphenicol. Expression of the VP22-Cre and Δ VP22-Cre fusion proteins could be induced by the addition of 0,5 mM IPTG to the culture medium. Analysis of protein extracts using Coomassie staining and

Western blotting of SDS-PAGE gels revealed that 50 - 60 % of VP22-Cre and Δ VP22-Cre proteins accumulated as insoluble inclusion bodies. The inclusion bodies where extracted and dissolved in 8 M urea. VP22-Cre and Δ VP22-Cre fusion proteins were subsequently purified using ion exchange chromatography. The quantity and purity of the isolated VP22-Cre and Δ VP22-Cre fusion proteins was determined using Coomassie stained SDS-PAGE gels and Western blot analysis (figure 8).

To analyse the ability of the purified fusion proteins to transduce into cultured cells, we used the fibroblast cell line M5Pax8 that contains a loxPcontaining reporter construct. When recombined by Cre recombinase, the reporter allows the expression of a β-galacosidase gene (Buchholz et al, Nucleic Acids Res. 24, 4256-4262, 1996). The cells where cultured for 18 h with increasing concentrations of VP22-Cre and Δ VP22-Cre in serum-free medium and analysed 4 days later for B-Galacosidase activity. Staining with X-Gal showed ~2 % blue cells in the cultures treated with up to 15 μg/ml ΔVP22-Cre indicating recombination of the loxP-flanked reporter construct had occurred. In contrast, cell cultures treated with up to 0,5 µg/ml VP22-Cre did not show any X-gal staining (figure 9). Measurement of cell lysates revealed a strong increase of β-galactosidase activity upon Δ VP22-Cre treatment when compared to untreated cells (figure 10). Genomic DNA was isolated fand subjected to PCR amplification that specifically amplifies a 250 bp fragment of the recombined allele. The deleted allele could be detected in cells treated with both VP22-Cre and Δ VP22-Cre fusion proteins (figure 11).

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This experiment demonstrates that VP22-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting.

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WO 01/49832 PCT/EP01/00060

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Gln Ala Phe Ser Glu His Thr Trp Lys Met Leu Leu Ser Val Cys Arg
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His Arg Arg Ser Gly Leu Pro Arg Pro Ser Asp Ser Asn Ala Val Ser 115 120 125

Leu Val Met Arg Arg Ile Arg Lys Glu Asn Val Asp Ala Gly Glu Arg 130 135 140

Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr Asp Phe Asp Gln Val Arg 145 150 155 160

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Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu Arg Ile Ala Glu Ile Ala 180 185 190

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Ile His Ile Gly Arg Thr Lys Thr Leu Val Ser Thr Ala Gly Val Glu 210 215 220

Lys Ala Leu Ser Leu Gly Val Thr Lys Leu Val Glu Arg Trp Ile Ser 225 230 235 240

Val Ser Gly Val Ala Asp Asp Pro Asn Asn Tyr Leu Phe Cys Arg Val 245 250 255

Arg Lys Asn Gly Val Ala Ala Pro Ser Ala Thr Ser Gln Leu Ser Thr 260 265 270

Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr His Arg Leu Ile Tyr Gly 275 280 285

Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu Ala Trp Ser Gly His Ser 290 295 300

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PCT/EP01/00060 WO 01/49832

29

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155

150

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gcg Ala 305	cct Pro	tat Tyr	cca Pro	atc Ile	ttt Phe 310	gct Ala	ata Ile	aag Lys	aat Asn	ggc Gly 315	cca Pro	aaa Lys	tct Ser	cac His	att Ile 320	960
		cat His														1008
ttg Leu	act Thr	aat Asn	gtt Val 340	gtg Val	gga Gly	aat Asn	tgg Trp	agc Ser 345	gat Asp	aag Lys	cgt Arg	gct Ala	tct Ser 350	gcc Ala	gtg Val	1056
		aca Thr 355														1104
ttc Phe	gca Ala 370	cta Leu	gtt Val	tct Ser	cgg Arg	tac Tyr 375	tat Tyr	gca Ala	tat Tyr	gat Asp	cca Pro 380	ata Ile	tca Ser	aag Lys	gaa Glu	1152
		gca Ala														1200
		cag Gln														1248

tgg aat ggg ata ata tca cag gag gta cta gac tac ctt tca tcc tac 1296
Trp Asn Gly Ile Ile Ser Gln Glu Val Leu Asp Tyr Leu Ser Ser Tyr
420 425 430

ata aat aga cgc ata taatga Ile Asn Arg Arg Ile 435

1317

<210> 4

<211> 437

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: DNA sequence
 coding for a fusion protein TAT-Flpe

<400> 4

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Gln Phe Asp Ile Leu Cys Lys Thr Pro Pro Lys Val Leu Val Arg Gln 20 25 30

Phe Val Glu Arg Phe Glu Arg Pro Ser Gly Glu Lys Ile Ala Ser Cys 35 40 45

Ala Ala Glu Leu Thr Tyr Leu Cys Trp Met Ile Thr His Asn Gly Thr 50 60

Ala Ile Lys Arg Ala Thr Phe Met Ser Tyr Asn Thr Ile Ile Ser Asn 65 70 75 80

Ser Leu Ser Phe Asp Ile Val Asn Lys Ser Leu Gln Phe Lys Tyr Lys 85 90 95

Thr Gln Lys Ala Thr Ile Leu Glu Ala Ser Leu Lys Lys Leu Ile Pro 100 105 110

Ala Trp Glu Phe Thr Ile Ile Pro Tyr Asn Gly Gln Lys His Gln Ser 115 120 125

Asp Ile Thr Asp Ile Val Ser Ser Leu Gln Leu Gln Phe Glu Ser Ser 130 135 140

Glu Glu Ala Asp Lys Gly Asn Ser His Ser Lys Lys Met Leu Lys Ala 145 150 155 160

Leu Leu Ser Glu Gly Glu Ser Ile Trp Glu Ile Thr Glu Lys Ile Leu 165 170 175

Asn Ser Phe Glu Tyr Thr Ser Arg Phe Thr Lys Thr Lys Thr Leu Tyr 180 - 185 190

Gln Phe Leu Phe Leu Ala Thr Phe Ile Asn Cys Gly Arg Phe Ser Asp 195 200 205

Ile Lys Asn Val Asp Pro Lys Ser Phe Lys Leu Val Gln Asn Lys Tyr 210 215 220

Leu Gly Val Ile Ile Gln Cys Leu Val Thr Glu Thr Lys Thr Ser Val 225 230 235 240 WO 01/49832 PCT/EP01/00060

32

Ser Arg His Ile Tyr Phe Phe Ser Ala Arg Gly Arg Ile Asp Pro Leu 245

Val Tyr Leu Asp Glu Phe Leu Arg Asn Ser Glu Pro Val Leu Lys Arg 260

Val Asn Arg Thr Gly Asn Ser Ser Ser Asn Lys Gln Glu Tyr Gln Leu 275 280 285

Leu Lys Asp Asn Leu Val Arg Ser Tyr Asn Lys Ala Leu Lys Lys Asn 290 295 300

Ala Pro Tyr Pro Ile Phe Ala Ile Lys Asn Gly Pro Lys Ser His Ile 305. 310 315 320

Gly Arg His Leu Met Thr Ser Phe Leu Ser Met Lys Gly Leu Thr Glu 325 330 335

Leu Thr Asn Val Val Gly Asn Trp Ser Asp Lys Arg Ala Ser Ala Val 340 345 350

Ala Arg Thr Thr Tyr Thr His Gln Ile Thr Ala Ile Pro Asp His Tyr 355 360 365

Phe Ala Leu Val Ser Arg Tyr Tyr Ala Tyr Asp Pro Ile Ser Lys Glu 370 380

Met Ile Ala Leu Lys Asp Glu Thr Asn Pro Ile Glu Glu Trp Gln His 385 390 395 400

Ile Glu Gln Leu Lys Gly Ser Ala Glu Gly Ser Ile Arg Tyr Pro Ala
405 410 415

Trp Asn Gly Ile Ile Ser Gln Glu Val Leu Asp Tyr Leu Ser Ser Tyr 420 425 430

Ile Asn Arg Arg Ile 435

<210> 5

<211> 2004

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence coding for a fusion protein VP22-Cre

<220>

<221> CDS

<222> (1)..(2001)

<400> 5

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1 5 10 - 15

gat gag tac gag gat ctg tac tac acc ccg tct tca ggt atg gcg agt 96
Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser
20 25 30

cc. Pr	c ga o As _l	t agt Sei 35	. FI	g cct	gad Asp	aco Thi	tcc Ser 40	Arc	cgt Arq	g Gly	gco Ala	c cta Leu 45	ı Glr	g aca	a cgc r Arg	144
t co Sei	g cgo r Aro	i ert	g ago	: g Gly	gag Glu	gto Val	. Arg	tto Phe	gto Val	caç L Glr	tad Ty:	c Asp	gag Glu	j tog Sei	g gat Asp	192
tat Ty:	. vr.	cto Lev	tac Tyr	: Gly	ggc Gly 70	Sex	tct Ser	tcc Ser	gaa Glu	gac Asp 75	Asp	gaa Glu	cac His	ccg Pro	g gag Glu 80	240
gto Val	e cco	cgg Arg	ace Thr	r cgg Arg 85	Arg	ccc	gtt Val	tcc Ser	ggg Gly 90	Ala	gtt Val	: ttg . Leu	tcc Ser	ggc Gly 95	ccg Pro	288
GJ À	r cct	gcg Ala	Arg	Ата	cct	ccg Pro	cca Pro	ccc Pro 105	gct Ala	ggg Gly	tcc Ser	gga Gly	ggg Gly	Ala	gga Gly	336
cgc Arg	aca Thr	CCC Pro 115	Thr	acc Thr	gec Ala	ccc Pro	egg Arg 120	gcc Ala	ccc Pro	cga Arg	acc Thr	cag Gln 125	cgg Arg	gtg Val	gcg Ala	384
act Thr	aag Lys 130	HΤα	ccc Pro	gcg Ala	gcc Ala	ccg Pro 135	gcg Ala	gcg Ala	gag Glu	acc Thr	acc Thr 140	cgc Arg	ggc Gly	agg Arg	aaa Lys	432
tcg Ser 145	Ara	cag Gln	cca Pro	gaa Glu	tcc Ser 150	gcc Ala	gca Ala	ctc Leu	cca Pro	gac Asp 155	gcc Ala	ccc Pro	gcg Ala	tcg Ser	acg Thr 160	480
gcg Ala	cca Pro	acc Thr	cga Arg	tcc Ser 165	aag Lys	aca Thr	ccc Pro	gcg Ala	cag Gln 170	ggg Gly	ctg Leu	gcc Ala	aga Arg	aag Lys 175	ctg . Leu	528
cac His	ttt Phe	agc Ser	acc Thr 180	gcc Ala	ccc Pro	cca Pro	aac Asn	ccc Pro 185	gac Asp	gcg Ala	cca Pro	tgg Trp	acc Thr 190	ccc Pro	cgg Arg	576
gtg Val	gcc Ala	ggc Gly 195	ttt Phe	aac Asn	aag Lys	cgc Arg	gtc Val 200	ttc Phe	tgc Cys	gcc Ala	gcg Ala	gtc Val 205	Gly aaa	cgc Arg	ctg Leu	624
gcg Ala	gcc Ala 210	atg Met	cat His	gcc Ala	Arg Arg	atg Met 215	gcg Ala	gcg Ala	gtc Val	cag Gln	ctc Leu 220	tgg Trp	gac Asp	atg Met	tcg Ser	67Ż
cgt Arg 225	ccg Pro	cgc Arg	aca Thr	gac Asp	gaa Glu 230	gac Asp	ctc Leu	aac Asn	gaa Glu	ctc Leu 235	ctt Leu	ggc Gly	atc Ile	acc Thr	acc Thr 240	720
atc Ile	cgc Arg	gtg Val	acg Thr	gtc Val 245	tgc Cys	gag Glu	ggc Gly	aaa Lys	aac Asn 250	ctg Leu	ctt Leu	cag Gln	cgc Arg	gcc Ala 255	aac Asn	768
gag Glu	ttg Leu	gtg Val	aat Asn 260	cca Pro	gac Asp	gtg Val	Val	cag Gln 265	gac Asp	gtc Val	gac Asp	Ala	gcc Ala 270	acg Thr	gcg Ala	816
act Thr	cga Arg	ggg Gly 275	cgt Arg	tct Ser	gcg Ala	АДа	tcg Ser 280	cgc Arg	ccc Pro	acc Thr	gag Glu	cga Arg 285	cct Pro	cga Arg	gcc Ala	864

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cca Pro	gcc Ala 290	Arg	tcc Ser	gct	tct Ser	cgc Arg 295	ccc Pro	aga Arg	cgg	ccc Pro	gtc Val 300	gag Glu	ggt Gly	acc Thr	gag Glu	912
ctc Leu 305	Gly	tcc Ser	act Thr	agt Ser	cca Pro 310	gtg Val	tgg Trp	tgg Trp	aat Asn	tct Ser 315	gca Ala	gat Asp	atc Ile	cag Gln	cac His 320	960
agt Ser	Gly	ggć Gly	cgc Arg	atg Met 325	Ser	aat Asn	tta Leu	ctg Leu	acc Thr 330	gta Val	cac His	caa Gln	aat Asn	ttg Leu 335	cct Pro	1008
gca Ala	tta Leu	ccg Pro	gtc Val 340	gat Asp	gca Ala	acg Thr	agt Ser	gat Asp 345	gag Glu	gtt Val	cgc Arg	aag Lys	aac Asn 350	ctg Leu	atg Met	1056
gac Asp	atg Met	ttc Phe 355	agg Arg	gat Asp	cgc Arg	cag Gln	gcg Ala 360	ttt Phe	tct Ser	gag Glu	cat His	acc Thr 365	tgg Trp	aaa Lys	atg Met	1104
ett Leu	etg Leu 370	ccc Ser	gtt Val	tğc Cys	cgg Arg	tcg Ser 375	tgg Trp	gcg Ala	gca Ala	tgg Trp	tgc Cys 380	aag Lys	ttg Leu	aat Asn	aac Asn	1152
cgg Arg 385	aaa Lys	tgg Trp	ttt Phe	ccc Pro	gca Ala 390	gaa Glu	cct Pro	gaa Glu	gat Asp	ġtt Val 395	cgc Arg	gat Asp	tat Tyr	ctt Leu	cta Leu 400	1200
tat Tyr	ctt Leu	cag Gln	gcg Ala	cgc Arg 405	ggt Gly	ctg Leu	gca Ala	gta Val	aaa Lys 410	act Thr	atc Ile	cag Gln	caa Gln	cat His 415	ttg Leu	1248
ggc Gly	cag Gln	cta Leu	aac Asn 420	atg Met	ctt Leu	cat His	cgt Arg	cgg Arg 425	tcc Ser	ggg Gly	ctg Leu	cca Pro	cga Arg 430	cca Pro	agt Ser	1296
gac Asp	agc Ser	aat Asn 435	gct Ala	gtt Val	tca Ser	ctg Leu	gtt Val 440	atg Met	cgg Arg	cgg Arg	atc Ile	cga Arg 445	aaa Lys	gaa Glu	aac Asn	1344
gtt Val	gat Asp 450	gcc Ala	ggt Gly	gaa Glu	cgt Arg	gca Ala 455	Lys	cag Gln	gct Ala-	cta Leu	gcg Ala 460	ttc Phe	gaa Glu	.cgc Arg	act Thr	1392
gat Asp 465	ttc Phe	gac Asp	cag Gln	gtt Val	cgt Arg 470	tca Ser	ctc Leu	atg Met	gaa Glu	aat Asn 475	agc Ser	gat Asp	cgc Arg	tgc Cys	cag Gln 480	1440
gat Asp	ata Ile	cgt Arg	aat Asn	ctg Leu 485	gca Ala	ttt Phe	ctg Leu	gly ggg	att Ile 490	gct Ala	tat Tyr	aac Asn	acc. Thr	ctg Leu 495	tta Leu	1488
cgt Arg	ata Ile	gcc Ala	gaa Glu 500	att Ile	gcc Ala	agg Arg	atc Ile	agg Arg 505	gtt Val	aaa Lys	gat Asp	atc Ile	tca Ser 510	cgt Arg	act Thr	1536
gac Asp	ggt Gly	ggg Gly 515	aga Arg	atg Met	tta Leu	atc Ile	cat His 520	att Ile	Gly ggc	aga Arg	acg [.] Thr	aaa Lys 525	acg Thr	ctg Leu	gtt Val	1584
agc Ser	acc Thr 530	gca Ala	ggt Gly	gta Val	gag Glu	aag Lys 535	gca Ala	ctt Leu	agc Ser	ctg Leu	ggg Gly 540	gta Val	act Thr	aaa Lys	ctg Leu	1632

U

gtc gag cga tgg att tcc gtc tct ggt gta gct gat gat ccg aat	
Val Glu Arg Trp Ile Ser Val Ser Gly Val Ala Asp Asp Pro Asn 545 550 555	aac 1680 Asn 560
tac ctg ttt tgc cgg gtc aga aaa aat ggt gtt gcc gcg cca tct Tyr Leu Phe Cys Arg Val Arg Lys Asn Gly Val Ala Ala Pro Ser 565 570 575	gcc 1728 Ala
acc agc cag cta tca act cgc gcc ctg gaa ggg att ttt gaa gca Thr Ser Gln Leu Ser Thr Arg Ala Leu Glu Gly Ile Phe Glu Ala 580 585 590	act 1776 Thr
cat cga ttg att tac ggc gct aag gat gac tct ggt cag aga tac His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr 595 600 605	ctg 1824 Leu
gcc tgg tct gga cac agt gcc cgt gtc gga gcc gcg cga gat atg Ala Trp Ser Gly His Ser Ala Arg Val Gly Ala Ala Arg Asp Met 610 615 620	gcc 1872 Ala
cgc gct gga gtt tea ata eeg gag atc atg caa gct ggt ggc tgg Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp 625 630 635	acc 1920 Thr 640
aat gta aat att gtc atg aac tat atc cgt aac ctg gat agt gaa Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu 645 650 655	aca 1968 Thr
ggg gca atg gtg cgc ctg ctg gaa gat ggc gat tag Gly Ala Met Val Arg Leu Leu Glu Asp Gly Asp 660 665	2004
<210> 6 <211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence coding for a fusion protein VP22-Cre	
<211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence	Arg
<211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence coding for a fusion protein VP22-Cre <400> 6 Met Thr Ser Arg Arg Ser Val Lys Ser Gly Pro Arg Glu Val Pro	
<pre><211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence</pre>	Ser
<pre><211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence</pre>	Ser Arg
<pre><211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence coding for a fusion protein VP22-Cre <400> 6 Met Thr Ser Arg Arg Ser Val Lys Ser Gly Pro Arg Glu Val Pro 1</pre>	Ser Arg Asp
<pre><211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence</pre>	Ser Arg Asp Glu 80
<pre><211> 667 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: DNA sequence</pre>	Ser Arg Asp Glu 80 Pro

17

Thr Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Thr Arg Gly Arg Lys Ser Ala Gln Pro Glu Ser Ala Ala Leu Pro Asp Ala Pro Ala Ser Thr 150 Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr 235 Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala 265 Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala 280 Pro Ala Arg Ser Ala Ser Arg Pro Arg Pro Val Glu Gly Thr Glu Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His Ser Gly Gly Arg Met Ser Asn Leu Leu Thr Val His Gln Asn Leu Pro 330 Ala Leu Pro Val Asp Ala Thr Ser Asp Glu Val Arg Lys Asn Leu Met Asp Met Phe Arg Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met 360 Leu Leu Ser Val Cys Arg Ser Trp Ala Ala Trp Cys Lys Leu Asn Asn Arg Lys Trp Phe Pro Ala Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu Tyr Leu Gln Ala Arg Gly Leu Ala Val Lys Thr Ile Gln Gln His Leu Gly Gln Leu Asn Met Leu His Arg Arg Ser Gly Leu Pro Arg Pro Ser 425 Asp Ser Asn Ala Val Ser Leu Val Met Arg Arg Ile Arg Lys Glu Asn Val Asp Ala Gly Glu Arg Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr

37

Asp Phe Asp Gln Val Arg Ser Leu Met Glu Asn Ser Asp Arg Cys Gln 465 470 475 480

Asp Ile Arg Asn Leu Ala Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu 485 490 495

Arg Ile Ala Glu Ile Ala Arg Ile Arg Val Lys Asp Ile Ser Arg Thr 500 : 505 510

Asp Gly Gly Arg Met Leu Ile His Ile Gly Arg Thr Lys Thr Leu Val 515 520 525

Ser Thr Ala Gly Val Glu Lys Ala Leu Ser Leu Gly Val Thr Lys Leu 530 535 540

Val Glu Arg Trp Ile Ser Val Ser Gly Val Ala Asp Asp Pro Asn Asn 545 550 555

Tyr Leu Phe Cys Arg Val Arg Lys Asn Gly Val Ala Ala Pro Ser Ala 565 570 575

Thr Ser Gln Leu Ser Thr Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr 580 585 590

His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu 595 600 605

Ala Trp Ser Gly His Ser Ala Arg Val Gly Ala Ala Arg Asp Met Ala 610 615 620

Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp Thr 625 630 635 .640

Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr 645 650 655

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<210> 7

17

<211> 2247

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence coding for a fusion protein VP22-Flpe

<220>

<221> CDS

<222> (1)..(2241)

<400> 7

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1 5 10 15

gat gag tac gag gat ctg tac tac acc ccg tct tca ggt atg gcg agt 96
Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser
20 . 25 . 30

ccc Pro	gat Asp	agt Ser 35	Pro	cct Pro	gac Asp	acc Thr	tcc Ser 40	cgc Arg	cgt Arg	ggc Gly	gcc Ala	cta Leu 45	cag Gln	aca Thr	cgc Arg	144
tcg Ser	cgc Arg 50	Gln	agg Arg	ggc Gly	gag Glu	gtc Val 55	cgt Arg	ttc Phe	gtc Val	cag Gln	tac Tyr 60	gac Asp	gag Glu	tcg Ser	gat Asp	192
tat Tyr 65	Ala	ctc Leu	tac Tyr	Gly	ggc Gly 70	tcg Ser	tct Ser	tcc Ser	gaa Glu	gac Asp 75	gac Asp	gaa Glu	cac His	ccg Pro	gag Glu 80	240
gtc Val	ccc Pro	cgg Arg	acg Thr	cgg Arg 85	cgt Arg	ccc Pro	gtt Val	tcc Ser	ggg Gly 90	gcg Ala	gtt Val	ttg Leu	tcc Ser	ggc Gly 95	ccg Pro	288
GJÀ āāā	cct Pro	gcg Ala	cgg Arg 100	gcg Ala	cct Pro	ccg Pro	cca Pro	ccc Pro 105	gct Ala	ggg	tcc Ser	gga Gly	ggg Gly 110	gcc Ala	gga Gly	336
cgc Arg	aca Thr	ccc Pro 115	acc Thr	acc Thr	gcc Ala	ccc Pro	cgg Arg 120	gcc Ala	ccc Pro	cga Arg	acc Thr	cag Gln 125	cgg Arg	gtg Val	gcg Ala	384
act Thr	aag Lys 130	gcc Ala	ccc Pro	gcg Ala	gcc Ala	ccg Pro 135	gcg Ala	gcg Ala	gag Glu	acc Thr	acc Thr 140	cgc Arg	ggc Gly	agg Arg	aaa Lys	432
tcg Ser 145	gcc Ala	cag Gln	cca Pro	gaa Glu	tcc Ser 150	gcc Ala	gca Ala	ctc Leu	cca Pro	gac Asp 155	gcc Ala	ccc Pro	gcg Ala	tcg Ser	acg Thr 160	480
gcg Ala	cca Pro	acc Thr	cga Arg	tcc Ser 165	aag Lys	aca Thr	ccc Pro	gcg Ala	cag Gln 170	ggg	ctg Leu	gcc Ala	aga Arg	aag Lys 175	ctg Leu	528
cac His	ttt Phe	agc Ser	acc Thr 180	gcc Ala	ccc Pro	cca Pro	aac Asn	ccc Pro 185	gac Asp	gcg Ala	cca Pro	tgg Trp	acc Thr 190	ccc Pro	cgg Arg	576
gtg Val	gcc Ala	ggc Gly 195	ttt Phe	aac Asn	aag Lys	cgc Arg	gtc Val 200	ttc Phe	tgc Cys	gcc Ala	gcg Ala	gtc Val 205	ggg Gly	cgc Arg	ctg Leu	624
gcg Ala	gcc Ala 210	atg Met	cat His	gcc Ala	cgg Arg	atg Met 215	gcg Ala	gcg Ala	gtc Val	cag Gln	ctc Leu 220	tgg Trp	gac Asp	atg Met	tcg Ser	672
 cgt Arg 225	ccg Pro	cgc Arg	aca Thr	gac Asp	gaa Glu 230	gac Asp	cte Leu	aac Asn	gaa Glu	ctc Leu 235	ctt Leu	ggc Gly	atc Ile	acc Thr	acc Thr 240	720
atc Ile	cgc Arg	gtg Val	acg Thr	gtc Val 245	tgc Cys	gag Glu	ggc Gly	aaa Lys	aac Asn 250	ctg Leu	ctt Leu	cag Gln	cgc Arg	gcc Ala 255	aac Asn	768
gag Glu	ttg Leu	gtg Val	aat Asn 260	cca Pro	gac Asp	gtg Val	gtg Val	cag Gln 265	gac Asp	gtc Val	gac Asp	gcg Ala	gcc Ala 270	acg Thr	gcg Ala	816
act Thr	cga Arg	ggg Gly 275	cgt Arg	tct Ser	gcg Ala	gcg Ala	tcg Ser 280	cgc Arg	ccc Pro	acc Thr	gag Glu	cga Arg 285	cct Pro	cga Arg	gcc Ala	864

cca Pro	gcc Ala 290	cgc Arg	tcc Ser	gct Ala	tct Ser	cgc Arg 295	ccc Pro	aga Arg	cgg Arg	ccc Pro	gtc Val 300	gag Glu	ggt Gly	acc Thr	gag Glu	912
ctc Leu 305	gga Gly	tcc Ser	act Thr	agt Ser	cca Pro 310	gtg Val	tgg Trp	tgg Trp	aat Asn	tct Ser 315	gca Ala	gat Asp	atc Ile	cag Gln	cac His 320	960
agt Ser	ggc	ggc Gly	cgc Arg	atg Met 325	agt Ser	caa Gln	ttt Phe	gat Asp	ata Ile 330	tta Leu	tgt Cys	aaa Lys	aca Thr	cca Pro 335	cct Pro	1008
aag Lys	gtc Val	ctg Leu	gtt Val 340	cgt Arg	cag Gln	ttt Phe	gtg Val	gaa Glu 345	agg Arg	ttt Phe	gaa Glu	aga Arg	cct Pro 350	tca Ser	GJÀ ààà	1056
					tgt Cys											1104
att Ile	act Thr 370	cat His	aac Asn	gga Gly	aca Thr	gca Ala 375	atc Ile	aag Lys	aga Arg	gcc Ala	aca Thr 380	ttc Phe	atg Met	agc Ser	tat Tyr	1152
aat Asn 385	act Thr	atc Ile	ata Ile	agc Ser	aat Asn 390	tcg Ser	ctg Leu	agt Ser	ttc Phe	gat Asp 395	att Ile	gtc Val	aac Asn	aaa Lys	tca Ser 400	1200
ctc Leu	cag Gln	ttt Phe	aaa Lys	tac Tyr 405	aag Lys	acg Thr	caa Gln	aaa Lys	gca Ala 410	aca Thr	att Ile	ctg Leu	gaa Glu	gcc Ala 415	tca Ser	. 1248
tta Leu	aag Lys	aaa Lys	tta Leu 420	att Ile	cct Pro	gct Ala	tgg Trp	gaa Glu 425	ttt Phe	aca Thr	att Ile	att Ile	cct Pro 430	tac Tyr	aat Asn	1296
gga Gly	caa Gln	aaa Lys 435	cat His	caa Gln	tct Ser	gat Asp	atc Ile 440	act Thr	gat Asp	att Ile	gta Val	agt Ser 445	agt Ser	ttg Leu	caa Gln	1344
					tcg Ser											1392
					gca Ala 470											1440
					cta Leu											1488
					tac Tyr											1536
					gat Asp											1584
					tat Tyr											1632

gag Glu 545	aca Thr	aag Lys	aca Thr	agc Ser	gtt Val 550	agt Ser	agg Arg	cac His	ata Ile	tac Tyr 555	ttc Phe	ttt Phe	agc Ser	gca Ala	agg Arg 560	1680
ggt Gly	agg Arg	atc Ile	gat Asp	cca Pro 565	ctt Leu	gta Val	tat Tyr	ttg Leu	gat Asp 570	gaa Glu	ttt Phe	ttg Leu	agg Arg	aat Asn 575	tct Ser	1728
gaa Glu	cca Pro	gtc Val	cta Leu 580	aaa Lys	cga Arg	gta Val	aat Asn	agg Arg 585	acc Thr	ggc Gly	aat Asn	tct Ser	tca Ser 590	agc Ser	aac Asn	1776
aaa Lys	cag Gln	gaa Glu 595	tac Tyr	caa Gln	tta Leu	tta Leu	aaa Lys 600	gat Asp	aac Asn	tta Leu	gtc Val	aga Arg 605	tcg Ser	tac Tyr	aac Asn	1824
aag Lys	gct Ala 610	ttg Leu	aag Lys	aaa Lys	aat Asn	gcg Ala 615	cct Pro	tat Tyr	cca Pro	atc Ile	ttt Phe 620	gct Ala	ata Ile	aag Lys	aat Asn	1872
ggc Gly 625	cca Pro	aaa Lys	tct Ser	cac His	att Ile 630	gga Gly	aga Arg	cat His	ttg Leu	atg Met 635	acc Thr	tca Ser	ttt Phe	ctg Leu	tca Ser 640	1920
atg Met	aag Lys	ggc Gly	cta Leu	acg Thr 645	gag Glu	ttg Leu	act Thr	aat Asn	gtt Val 650	gtg Val	gga Gly	aat Asn	tgg Trp	agc Ser 655	gat Asp	1968
aag Lys	cgt Arg	gct Ala	tct Ser 660	gcc Ala	gtg Val	gcc Ala	agg Arg	aca Thr 665	acg Thr	tat Tyr	act Thr	cat His	cag Gln 670	ata Ile	aca Thr	2016
gca Ala	ata Ile	cct Pro 675	gat Asp	cac His	tac Tyr	ttc Phe	gca Ala 680	Leu	gtt Val	tct Ser	cgg Arg	tac Tyr 685	tat Tyr	gca Ala	tat Tyr	2064
gat Asp	cca Pro 690	ata Ile	tca Ser	aag Lys	gaa Glu	atg Met 695	ata Ile	gca Ala	ttg Leu	aag Lys	gat Asp 700	gag Glu	act Thr	aat Asn	cca Pro	2112
att Ile 705	gag Glu	gag Glu	tgg Trp	cag Gln	cat His 710	ata Ile	gaa Glu	cag Gln	cta Leu	aag Lys 715	ggt Gly	agt Ser	gct Ala	gaa Glu	gga Gly 720	2160
agc Ser	ata Ile	cga Arg	tac Tyr	ccc Pro 725	gca Ala	tgg Trp	aat Asn	Gly	ata Ile 730	Ile	tca Ser	cag Gln	gag Glu	gta Val 735	cta Leu	2208
gac Asp	tac Tyr	ctt Leu	tca Ser 740	tcc Ser	tac Tyr	ata Ile	aat Asn	aga Arg 745	Arg	ata Ile	taa	tga				2247

<210> 8

<211> 747

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: DNA sequence coding for a fusion protein VP22-Flpe

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Val	Ala	Gly 195	Phe	Asn	Lys	Arg	Val 200	Phe	Cys	Ala	Ala	Val 205	Gly	Arg	Leu
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gag ttg gtg aat cca gac gtg gtg cag gac gtc gac gcg gcc acg gcg 38 Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala 115 120 125	4
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tat Tyr	ctt Leu	cag Gln	gcg Ala 260	cgc Arg	ggt Gly	ctg Leu	gca Ala	gta Val 265	aaa Lys	act Thr	atc Ile	cag Gln	caa Gln 270	cat His	ttg Leu	816
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	agc Ser 290															912
	gat Asp															960
	ttc Phe															1008
	ata Ile															1056
	ata Ile															1104
	ggt Gly 370															1152
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	agc Ser															1344
	cga Arg 450															1392
	tgg Trp															1440

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PCT/EP01/00060

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	gcc Ala															624
	gcc Ala 210															672
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	gcc Ala 290															912
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				tgc Cys												1152
				ccc Pro												1200
				cgc Arg 405												1248
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Ser Arg Gln Arg Gly Glu Val Arg Phe Val Gln Tyr Asp Glu Ser Asp .50 55 60

Tyr Ala Leu Tyr Gly Gly Ser Ser Ser Glu Asp Asp Glu His Pro Glu 65 70 75 80

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Gly Pro Ala Arg Ala Pro Pro Pro Pro Ala Gly Ser Gly Gly Ala Gly
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Arg Thr Pro Thr Thr Ala Pro Arg Ala Pro Arg Thr Gln Arg Val Ala 115 120 125

Ser Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Thr Arg Gly Arg Lys 130 135 140

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WO 01/49832 PCT/EP01/00060

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His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu 595 600 605

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PCT/EP01/00060

21

WO 01/49832

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Claims

- 1. Use of a fusion protein comprising
- (a) a site-specific DNA recombinase domain and
- (b) a protein transduction domain (PTD)

for preparing an agent for inducing target gene alterations in a living organism or cell culture, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in an endogenous gene.

- 2. The use of claim 1, wherein the PTD is not derived from Antennapedia and preferably is a PTD derived from the VP22 protein of HSV or from the TAT protein of HIV.
- 3. Use of a fusion protein comprising
- (a) a site-specific DNA recombinase domain and
- (b) a protein transduction domain (PTD) being not derived from Antennapedia and preferably being derived from the VP22 protein of HSV or from the TAT protein of HIV

for preparing an agent for inducing target gene alterations in a living organism or cell culture, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome.

- 4. The use of claim 3, wherein the recognition sites for said site specific recombinase is present within an endogenous gene or a transgene.
- 5. The use of any one of claims 2 to 4, wherein the TAT protein comprises
- (i) the amino acid sequence YGRKKRRQRRR (SEQ ID NO: 10) or a mutant thereof including
- (ii) peptides having the amino sequences

71

AGRKKRRQRRR (SEQ ID NO:22)

YARKARRQARR (SEQ ID NO:23)

YARAAARQARA (SEQ ID NO:24)

YARAARRAARR (SEQ ID NO:25)

YARAARRAARA (SEQ ID NO:26)

YARRRRRRRR (SEQ ID NO:27)

YAAARRRRRRR (SEQ ID NO:28);

preferably the TAT protein consists of one of the sequences shown in (i) or (ii) above.

- 6. The use of any one of claims 2 to 4, wherein the VP22 protein comprises the amino acid 16-157 of SEQ ID NO:14.
- 7. The use of any one of claims 1 to 6, wherein the site-specific DNA recombinase domain is selected from a recombinase protein derived from Cre, Flp, ϕ C31 recombinase, and R recombinase and preferably is Cre having amino acids 15 to 357 of SEQ ID NO: 2 or Flpe having amino acids 15 to 437 of SEQ ID NO: 4.
- 8. The use of any one of claims 1 to 7, wherein the protein transduction domain is fused to the N-terminal of the site-specific DNA recombinase domain.
- 9. The use of any one of claims 1 to 8, wherein the protein transduction domain is fused to the site-specific DNA recombinase domain through a direct chemical bond or through a linker molecule.
- 10. The use of any one of claim 9, wherein the linker molecule is a short peptide having 1 to 20, preferably 1 to 10 amino acid residues.

WO 01/49832

11. The use of any one of claims 1 to 10, wherein said fusion protein further comprises additional functional sequences.

72

- 12. The use of claim 1, wherein the fusion protein has the sequence shown in SEQ ID NOs: 2, 4, 6 or 8.
- 13. The use of any one of claims 1 to 12, wherein the living organism is a vertebrate, preferably a rodent or a fish.
- 14. A method for inducing gene alterations in a living organism which comprises administering to said living organism, a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain as defined in claims 1 to 12, wherein said living organism carries at least one or more recognition sites for said sitespecific DNA recombinase integrated in its genome.
- 15. A fusion protein comprising
- (a) a site-specific DNA recombinase domain as defined in claims 2 to 9 and
- (b) a protein transduction domain (PTD) as defined in claims 2 to 9 provided that when (a) is the wild-type Flp or Cre then (b) is not the full length VP22 protein of HSV.
- 16. The fusion of claim 15, wherein the (PTD) is derived from the TAT protein of HIV.
- 17. A DNA sequence coding for the fusion protein of claim 15 or 16, said DNA sequence preferably comprising the sequence shown in SEQ ID NOs:1, 3, 5, 7, 9, 11, 13, 18 and/or 20.
- . 18. A vector comprising the DNA sequence of claim 17.

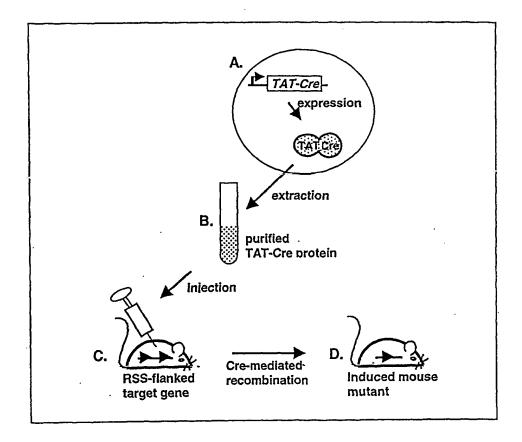
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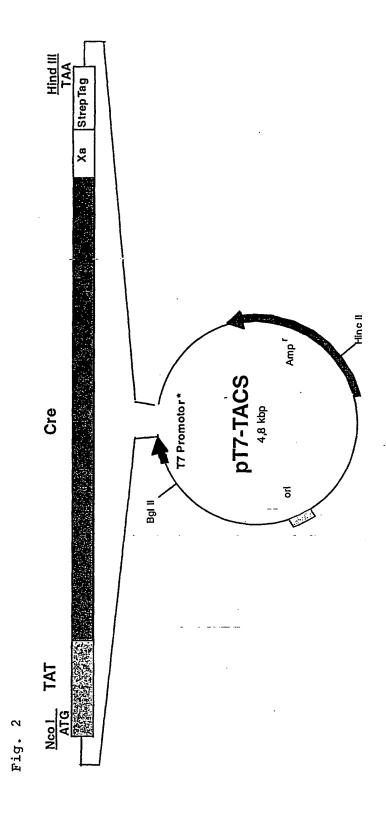
73

- 19. A host cell transformed with the vector of claim 18 and/or comprising the DNA of claim 17.
- 20. A method for producing the fusion protein of claim 15 which comprises culturing the transformed host cell of claim 19 and isolating the fusion protein.
- 21. An injectable composition comprising the fusion protein as defined in claims 1 to 12 or 15 to 16.

WO 01/49832 PCT/EP01/00060

Fig. 1





SUBSTITUTE SHEET (RULE 26)

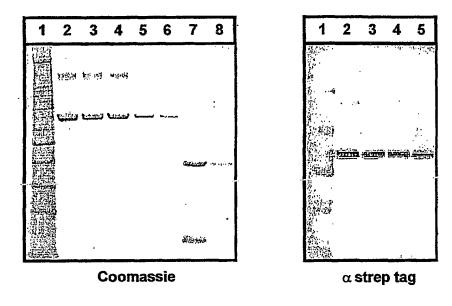


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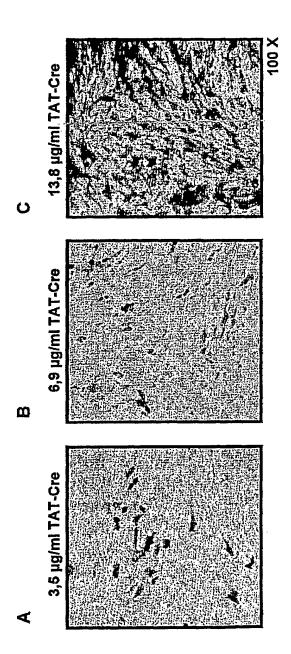
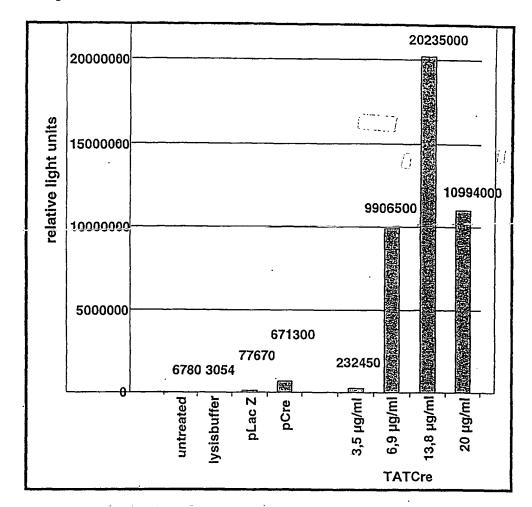


Figure 4

Fig. 5



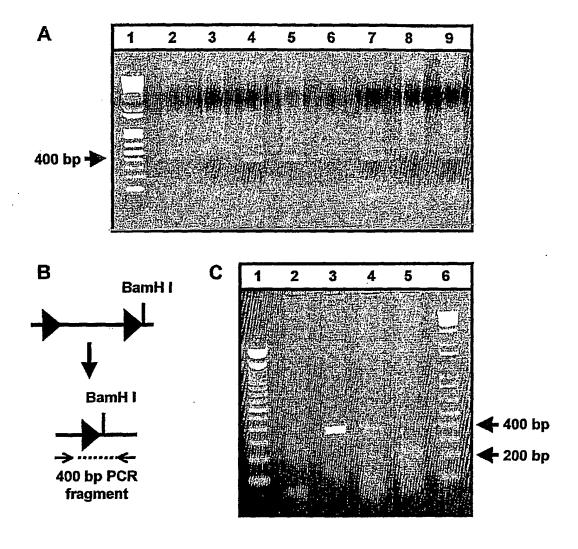
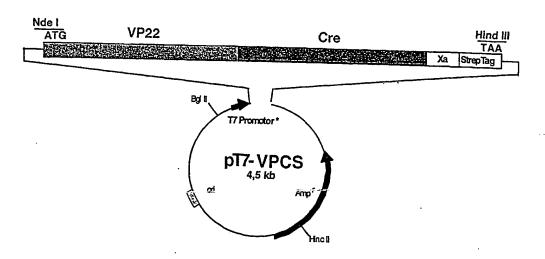
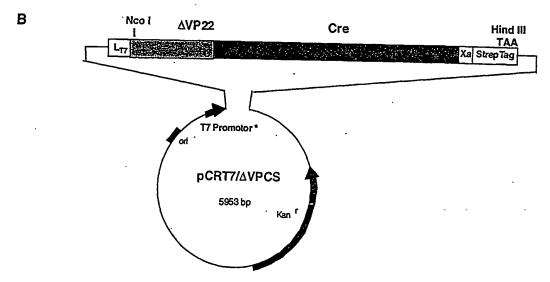


Figure 6

Fig. 7







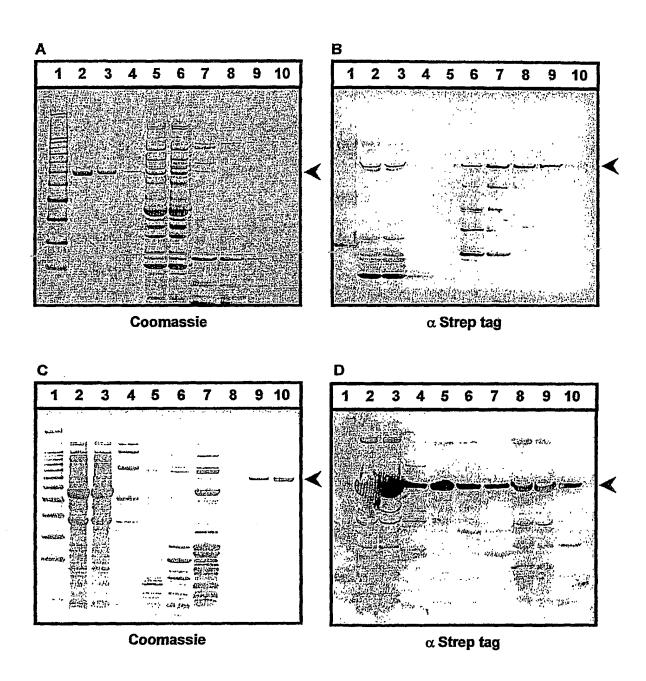


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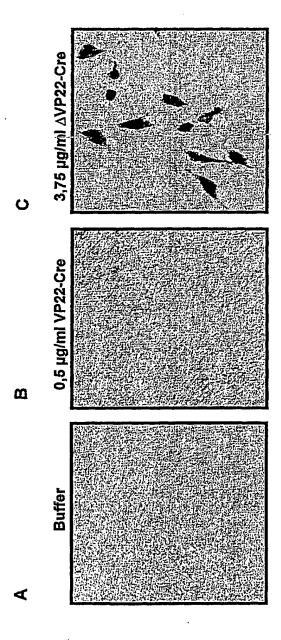
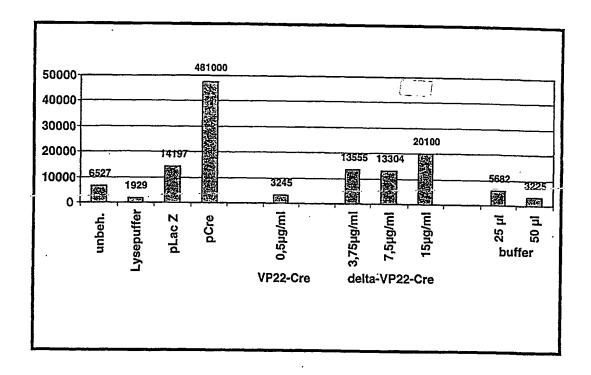


Figure 9

Fig. 10



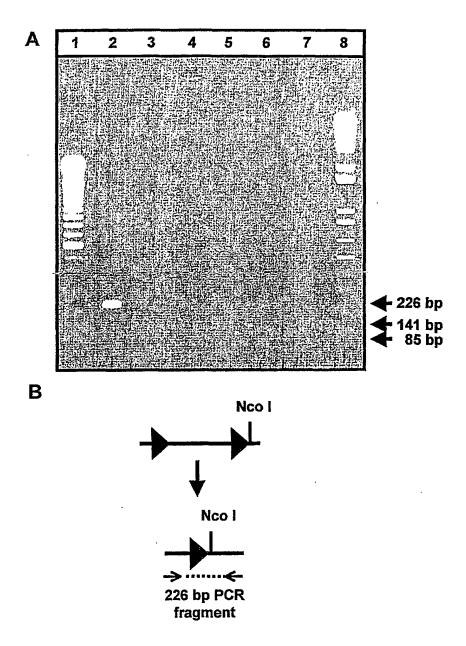


Figure 11